

### **Larval RNA in situ (from Peter Follette)**

- Fix 15'-20' in 4% formaldehyde in PBS
- Fix 15'-20' in 4% formaldehyde in PBT
- Wash 3 x 5' in PBT
- Proteinase K (10microg/ml) 3'-5'
- Wash 2 x 5' in 2mg/ml glycine in PBT
- Fix 15' in 4% formaldehyde in PBS
- Wash 5 x 5' in PBT
- Wash 10' in hybe solution (same solution as for embryos; see below)
- Prehybe at least 1hr @ 70
- Hybe (40ng riboprobe/100microliter) 24-36hrs @ 70
- Wash 20' @ 70 in hybe sol'n
- Wash 20' @ 70 in 1:1 hybe:PBT
- Wash 10-12hr in PBT @ 70, changing solutions 5 times
- Add 1:2000 anti-DIG-AP 1hr
- Wash 4 x 30' in PBT
- Wash 20' in staining buffer (same as for embryos, but no Triton X-100)
- Add 4.5 microliters NBT, 3.5 microliters X-phosphate per 1ml staining buffer

#### **Hybe solution:**

- 50% deionized formamide
- 5X SSC
- 100 ug/ml sonicated, denatured salmon sperm DNA
- 100 ug/ml E. coli tRNA
- 50 ug/ml heparin
- 0.1% Tween 20
- pH to 4.5 with Citric Acid

#### **Staining Buffer:**

- 100 mM Tris pH 9.5
- 100 mM NaCl
- 50 mM MgCl<sub>2</sub>